Review Article

Molecular signatures of epithelial-mesenchymal transition and their role in cancer progression

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Abstract

The differentiation of epithelial cells to mesenchymal cells is known as epithelial-mesenchymal transition (EMT) which plays a significant role in embryo and organ development, wound healing and tissue regeneration. Off late, EMT has emerged as a major factor contributing towards cancer metastasis. Cancer metastasis is a key challenge faced during the treatment of cancer. Apart from promoting cancer, EMT is also considered as a factor helping in the generation of cancer stem cells which can eventually lead to drug resistance making EMT a potential target in anticancer therapies. In this review, we highlight the different types of EMT, biomarkers of EMT which could be targeted to halt the EMT process which may provide new hope in cancer treatment.

Keywords: Epithelial-mesenchymal transition, cancer, biomarkers, metastasis

Introduction

Epithelial-mesenchymal transition (EMT) is a phenomenon in which a polarized epithelial cell, undergoes biochemical changes transforming it into a mesenchymal cell phenotype, resulting in invasiveness, increased migration, hoisted protection from apoptosis, improved generation of extracellular matrix (ECM) components (Kalluri and Nelson, 2003). EMT is initiated through some molecular processes which include activation of transcription factors, expression of specific cellsurface proteins, cytoskeletal proteins, production of ECMdegrading enzymes, and changes in the expression of specific microRNAs. At the end of an EMT process, the basement membrane undergoes degradation, and mesenchymal cells are formed which can migrate from their origin (Kalluri and Weinberg, 2009). The inverse of EMT is mesenchymalepithelial transition (MET) in which the migratory ability of the cells is lost, with cells depicting hallmarks of epithelial tissues namely undergoing apicobasal polarization and expressing the junctional complexes (Thiery et al., 2009). The process of EMT

was first described by Elizabeth Hay who observed EMT transition in the primitive streak of chick embryos (Hay, 1995). EMT is an important developmental process, usually activated in wound healing, fibrosis, and cancer metastasis (Thiery et al., 2009; Kalluri and Weinberg, 2009; Chapman, 2011). Differentiation of embryonic stem cells, behavior of cancer stem cells and induced pluripotency are also regulated by EMT and MET (Lamouille et al., 2014).

Epithelial and mesenchymal cells differ phenotypically as well as in function, although both types of cells have inherent plasticity in common (Lamouille et al., 2014). Epithelial cells are linked together by gap junctions, tight junctions, adherens and also have an apicobasal polarity, the polarization of the actin cytoskeleton and are joined by a basal lamina at their basal surface. Mesenchymal cells, on the other hand, do not have the polarity, morphologically are spindle-shaped and interact with each other through focal points (Thiery and Sleeman, 2006). Epithelial cells express high levels of E cadherin, while mesenchymal cells express N cadherin, vimentin, and fibronectin. Thus, EMT involves significant morphological as well as phenotypical changes to a cell.

Biologically, EMTs are of three types: Type I or developmental, Type II or fibrosis (Phua et al., 2013) and wound healing, and Type III or cancer metastasis (Li and Li, 2015).

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EMT classification

EMT is now being studied vastly to understand its role in different signaling and transcription pathways (Kalluri and Zeisberg, 2006). Thus, a proposal to classify EMTs into three different biological subtypes based on their occurrence was considered in a meeting on EMT in Poland (2007) followed by a meeting in March 2008 at Cold Spring Harbor Laboratories (Kalluri and Weinberg, 2009).

Type 1 EMT: During implantation, embryo and organ development

Type 1 EMT involves primitive epithelial cells transitioning to motile mesenchymal cells as part of gastrulation and primitive neuroepithelial cells generating migrating neural crest cells. In both situations, some of the cells generated by EMT are reinduced as secondary epithelial cells in mesodermal and endodermal organs by MET (Zeisberg and Neilson, 2009). Further, following the earliest stages of embryogenesis, embryo implantation and the initiation of the formation of the placenta are both associated with an EMT that involves the parietal endoderm (Vicovac and Aplin, 1996). In particular, the trophectoderm cells, which are precursors of the cytotrophoblast, undergo an EMT to facilitate the invasion of the endometrium and the subsequent proper anchoring of the placenta, enabling its function in nutrient and gas exchange (Bischof et al., 2006). At the biochemical level, the EMT associated with gastrulation is dependent on and orchestrated by canonical Wnt signaling, and embryos deficient in Wnt3 cannot undergo the EMT associated with gastrulation (Skromne and Stern, 2001). Subsequently, the formation of a primitive streak is related to the expression of Wnt8c, and ectopic expression of Wnt8c in embryos that leads to multiple primitive streaks (Thomas et al., 1997).

During embryonic development, an EMT involving the epithelial cells of the neuroectoderm gives rise to migratory neural crest cells (Duband and Thiery, 1982). Initially, the premigratory neural crest cells express genes such as *Sox, Snail, Slug, and forkhead box D3 (FoxD3)*, and these cells subsequently undergo an EMT (Liem et al., 2000). As a consequence, they then dissociate from the neural folds, become motile, and disperse to the different parts of the embryo, where they undergo further differentiation into other cell types (Kalluri and Weinberg, 2009).

Type 2 EMT: Related with regeneration of tissues and fibrosis of organ

EMT-Type 2 is associated with wound recovery, tissue regeneration, and organ fibrosis. In various epithelial tissues, organ fibrosis arises by the inflammatory cells and by fibroblast that induces the numerous inflammatory signals of ECM complex. This type of EMT is usually seen in kidney, lung, liver

and intestine fibrosis (Potenta et al., 2008; Zeisberg et al., 2007; Zeisberg et al., 2007; Kim et al., 2006). Furthermore essential tumor nodules of epithelial origin have cancerrelated fibroblasts that acquire certain types of genetic alteration /mutation with a tumor cell, thus leading to tumorigenesis (Yu et al., 2014).

Type 3 EMT: Related with the progression of cancer and metastasis

During cancer progression, cancer cells invade adjacent tissues and migrate from their primary site to distant sites causing cancer metastasis (Hanahan and Weinberg, 2000). In many cases, EMT has been observed to be a part of one of the many causes of cancer metastasis (Thiery 2002). Many mesenchymal markers such as S100A4 (also termed as FSP1 or fibroblast-specific protein-1) or vimentin (Thompson et al., 1994), nuclear overexpression of β -catenin (Brabletz et al., 1998) and E-cadherin (loss of epithelial adhesion molecule) are the biomarkers for cancer metastasis (Mareel et al., 1989). Type 3 EMT is essential for the transition and preparation of epithelial nodular tumor cells for metastasis, movement, invasion but not for the formation of fibroblasts (Xue et al., 2003).

Complete knowledge of signaling events contributing towards EMT is still unclear. One of the proposed ideas is that the cancer cells undergoing genetic and epigenetic changes during the formation of primary tumor make them responsive to EMT-inducing heterotypic changes which originate in the tumor-associated stroma (Smit and Peeper, 2008). EMT-inducing signals arising from PDGF (Plateletderived growth factor), EGF (Epidermal growth factor), HGF (Hepatocyte growth factor), and TGF-β (Transforming growth factor beta), play an essential role in activating EMT inducing transcription factors like Snail, Slug, Twist, Goosecoid, zinc finger E-box binding homeobox 1 (ZEB1) and FOXC2 (Forkhead box protein C2) in cancer cells and thus helping in cancer metastasis (Thiery, 2002; Jechlinger et al., 2002; Niessen et al., 2008; Medici et al., 2008). These transcription factors then help in activating the EMT program in cancer cells. A successful execution of EMT in cancer cells also depends on different signaling networks and signal transducing proteins such as PI3K, Akt, MAPK, ERK, Smads, β-catenin, lymphoid enhancer binding factor (LEF), RhoB, Ras, and c-Fos β4 integrins, α5β1 integrin, and αVβ6 integrin (Tse and Kalluri, 2007). Any disruption of cell-cell adherens junctions also promotes EMT process and the cell-ECM adhesions mediated by integrins facilitated by the disruption of cell-cell adheren junctions and the cell-ECM adhesions mediated by integrins (Gupta et al., 2005; Mani

et al., 2007; Taki et al., 2006). Therefore, EMT plays an essential role in cancer metastasis and thus helping in the formation of secondary tumors.

Role of Tumor Microenvironment in EMT

The tumor microenvironment plays a fundamental role in tumor progression, induction of EMT, and cancer metastasis. There are several cells in the tumor microenvironment, such as inflammatory and immune cells, cancer-associated fibroblasts (CAFs), extracellular matrix components (ECM), endothelial and epithelial cells, mesenchymal stem cells, etc. (Tse and Kalluri, 2007). Immune cells induce EMT by infiltrating the primary tumor and activating TFG-β, EGF, and HGF from several signaling pathways. Two signaling pathways namely TFG-β/Smad and NF-kB cause changes in mesenchymal phenotype and initiates the process of metastasis in vivo. Thus, targeting the NF-kB signaling pathway in cancer may protect against cancer cell metastasis (Labelle et al., 2011). Positive regulation of EMT markers such as N-cadherin, Vimentin, Twist and Snail and negative regulation of E-cadherin expression was noticed in co-culture of breast tumor cells with bone marrowderived mesenchymal stem cells (MSCs). Therefore, these cells can promote breast cancer metastasis by facilitating the EMT process (Martin et al., 2010). Tumor-associated macrophages (TAM) can also induce EMT in the intra-tumor environment by the activation of the β -catenin pathway and signaling of TFG- β . TAM can also induce EMT in the intra-tumor environment by the activation of the β-catenin pathway and signaling of TFG-β. A study conducted by Bonde et al. showed a positive correlation between TAM density and expression of mesenchymal markers, activation of \beta-catenin pathway, increase in mesenchymal markers expression, decrease in E-cadherin expression, and an invasive phenotype (Bonde et al., 2012). Thus, discerning the markers that help in the induction of EMT may help in thoroughly understanding the complicated process of EMT and its relation with tumor-microenvironment.

Biomarkers for EMT

EMT biomarkers are widely used to characterize the cancer cells. Some of the most common biomarkers used in EMT studies are TFG- β , Wnts, SNAIL and TWIST, cadherins and vimentin (Thompson et al., 2005). TGF- β is one of the most important inducers of EMT and a vital suppressor of epithelial cells (Pickup et al., 2013). TGF- β triggers cells to lose epithelial markers, like E-cadherin and helps them in gaining mesenchymal markers, such as vimentin. Wild-type TGF- β helps in cell proliferation but mutated TGF- β helps in the uncontrolled proliferation of cancer cells (Bellam and Pasche, 2010). Another transcription factor which is induced by TGF- β is Snail. It controls the protein expression of mesenchymal phenotype cells and also suppresses epithelial proteins like E-cadherin (Lamouille et al., 2014). Snail

also helps in promoting epithelial cell migration and differentiation while in the embryonic stage it promotes the formation of mesoderm (Fidler and Poste, 2008). Another transcription factor namely Twist helps in the migration and differentiation of epithelial to mesenchymal cells. Twist also promotes the conversion of E-cadherin to N- cadherin. N-cadherin is one of the important biomarkers of mesenchymal cells (Vaittinen et al., 2001). The expression of E-cadherins and N-cadherins are usually checked to screen the presence of EMT and to determine the presence of mesenchymal stage of cancer cells (Strutuz et al., 2002). Studies related to breast cancer have shown that the level of E-cadherin is inversely proportional to metastasis and poor prognosis. Thus, E-cadherin level is monitored to determine tumor progression (Singhai et al., 2011). Expression of Ecadherin is also affected by Wnt signaling pathway whose function is to control the transcription of genes that helps in cell proliferation, differentiation, and migration (Wang and Zhou, 2011). In EMT process, epithelial cells acquire mesenchymal phenotype and start expressing mesenchymal markers like vimentin, tyrosine kinase 2 receptor discoidin domain (DDR2), fibroblast-specific protein 1 (S100A4) and collagen-specific tyrosine kinase receptor (Ren et al., 2014). Vimentin is an intermediate filament, and upregulation in its expression has been observed in several cancer types such as breast, colon, prostate. Vimentin expression in these cancers is associated with metastasis and poor prognosis (Kalluri and Weinberg, 2009; Lehtinen et al., 2013). Thus, these results show that the expression of vimentin can be used as a biomarker to predict invasiveness and survival in breast cancer (Patel et al., 2015).

Overcoming Drug Resistance by Targeting EMT

EMT plays a vital role in drug resistance. In a study conducted by Gupta et al., EMT cells were generated by Ecadherin shRNA, and these cells were further used to identify the CSC-selective small molecule inhibitors. After screening, it was observed that salinomycin, an antibiotic can selectively kill breast CSCs (Gupta et al., 2009). It has also been observed that salinomycin can also inhibit EMT induced by doxorubicin treatment and further sensitize HCC cells towards doxorubicin (Zhou et al., 2015). Apart from salinomycin, other small molecule inhibitors of EMT have also been tested in-vitro and in-vivo. Curcumin was able to sensitize colorectal cells that were initially resistant to 5-fluorouracil via miRNA-mediated suppression of EMT (Toden et al., 2015). Mocetinostat, a histone deacetylase (HDAC) inhibitor, inhibited the expression of EMT transcription factor namely ZEB1 by restoring miR-203, changed the EMT phenotype in pancreatic cancer and sensitized cells towards docetaxel (Meidhof et al., 2015). It has also been reported that Akt/GSK3β/Snail1 pathway-driven EMT causes gemcitabine resistance in pancreatic cancer. Further, the sensitivity of gemcitabine towards pancreatic cancer cells was restored by the administration of zidovudine, an antiviral drug, which led to inhibition of the Akt/GSK3\beta/Snail1 signaling pathway. Tumor formation in mice having gemcitabine-resistant pancreatic tumor xenograft was inhibited when zidovudine with gemcitabine was co-administered and hence, halted the pancreatic cancer cells from acquiring EMT phenotype (Namba et al., 2015). Metformin an anti-diabetic drug has also been shown to inhibit cancer and is now being investigated as a potential anti-cancer agent (Evans et al., 2005). Hirsch et al. (2009) showed that metformin could selectively target breast cancer stem cells (BCSCs). Vazquez-Martin et al. (2010) reported that metformin downregulated the expression of important EMT transcription factors such as ZEB1, Twist1, and SNAI2 and induced transcriptional reprogramming of BCSCs (Vazquez-Martin et al., 2010). Metformin also blocked the IL-6/STAT3 pathway and further inhibited the lung cancer cells from acquiring mesenchymal phenotype (Zhao et al., 2014). The exact mechanism of action of metformin by which it inhibits EMT is still unclear, but it is speculated that metformin may work through AMPK activation and inhibits EMT formation (Chou et al., 2014). Thus metformin can become a suitable candidate as an anti-cancer compound possessing EMT inhibiting potential (Lv and Shim, 2015).

Conclusion

It is now strongly suggested that EMT plays a significant role in cancer progression by targeting multiple transcription factors which in turn can activate different pathways. EMT is becoming a potential target to treat cancer, and many studies suggest that targeting EMT can help in halting cancer metastasis and this provides a new ray of hope in cancer treatment.

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Conflict of interest

The authors declare no conflict of interest

References

- Behrens J, Mareel MM, Van Roy FM, Birchmeier W. 1989. Dissecting tumor cell invasion: epithelial cells acquire invasive properties after the loss of uvomorulin-mediated cell-cell adhesion. Journal of Cell Biology, 108(6):2435–2447.
- Bischof P, Aplin JD, Bentin-Ley U, Brannstrom M, Casslen B,

- Castrillo JL, Classen-Linke I, Critchley HO, Devoto L, D'Hooghe T, Horcajadas JA, Groothuis P, Ivell R, Pongrantz I, Macklon NS, Sharkey A, Vicovac L, White JO, Winterhager E, von Wolff M, Simon C, Stavreus-Evers A. 2006. Implantation of the human embryo: research lines and models. From the implantation research network 'Fruitful'. Gynecologic and Obstetric Investigation, 62(4):206–216.
- Bonde AK, Tischler V, Kumar S, Soltermann A, Schwendener RA. 2012. Intratumoral macrophages contribute to epithelial-mesenchymal transition in solid tumors. BMC Cancer, 24:12-35.
- Brabletz T, Jung A, Hermann K, Günther K, Hohenberger W, Kirchner T. 1998. Nuclear overexpression of the oncoprotein beta-catenin in colorectal cancer is localized predominantly at the invasion front. Pathology, Research and Practice, 194(10):701-704.
- Chapman HA. 2011. Epithelial-mesenchymal interactions in pulmonary fibrosis. Annual Review of Physiology, 73:413–435.
- Chou CC, Lee KH, Lai IL, Wang D, Mo X, Kulp SK, Shapiro CL, Chen CS. 2014. AMPK reverses the mesenchymal phenotype of cancer cells by targeting the Akt-MDM2-Foxo3a signaling axis. Cancer Research, 74(17):4783–4795.
- Duband JL, Thiery JP. 1982. Appearance and distribution of fibronectin during chick embryo gastrulation and neurulation. Developmental Biology, 94(2):337-350.
- Evans, JM, Donnelly LA, Emslie-Smith AM, Alessi DR, Morris AD. 2005. Metformin and reduced risk of cancer in diabetic patients. BMJ, 330(7503):1304–1305.
- Fidler IJ, Poste G. 2008. The "seed and soil" hypothesis revisited. Lancet Oncology, 9(8):808.
- Gupta PB, Mani S, Yang J, Hartwell K, Weinberg RA. 2005. The evolving portrait of cancer metastasis Cold Spring Harbor Symposia on Quantitative Biology, 70:291–297.
- Gupta PB, Onder TT, Jiang G, Tao K, Kuperwasser C, Weinberg RA, Lander ES. 2009. Identification of selective inhibitors of cancer stem cells by high-throughput screening. Cell 138(4):645–659.
- Hanahan D, Weinberg RA. 2000. The hallmarks of cancer. Cell. 100(1): 57–70.
- Hay ED. (1995). An overview of epithelio-mesenchymal transformation. Acta Anatomica 154(1): 8–20.
- Hirsch HA, Iliopoulos D, Tsichlis PN, Struhl K. 2009. Metformin selectively targets cancer stem cells and acts

- together with chemotherapy to block tumor growth and prolong remission. Cancer Research, 69(19):7507–7511.
- Jechlinger M, Grunert S, Beug H. 2002. Mechanisms in epithelial plasticity and metastasis: insights from 3D cultures and expression profiling. Journal of Mammary Gland Biology and Neoplasia, 7(4): 415–432.
- Kalluri R, Neilson EG. 2003. Epithelial mesenchymal transition and its implications for fibrosis. Journal of Clinical Investigation, 112(12):1776-1784.
- Kalluri R, Weinberg RA. 2009. The basics of epithelialmesenchymal transition. Journal of Clinical Investigation, 119(6):1420-1428.
- Kalluri R, Zeisberg M. 2006. Fibroblasts in cancer. Nature Reviews Molecular Cell Biology, 6(5):392–401.
- Kim KK, Kugler MC, Wolters PJ, Robillard L, Galvez MG, Brumwell AN, Sheppard D, Chapman HA. 2006. Alveolar epithelial cell mesenchymal transition develops in vivo during pulmonary fibrosis and is regulated by the extracellular matrix. Proc Proceedings of the National Academy of Sciences of the United States of America, 103(35):13180-13185.
- Labelle M, Begum S, Hynes RO. 2011. Direct signaling between platelets and cancer cells induces an epithelial-mesenchymal-like transition and promotes metastasis. Cancer Cell, 20(5):576-590.
- Lamouille S, Xu J, Derynck R. 2014. Molecular mechanisms of epithelial-mesenchymal transition. Nature Reviews Molecular Cell Biology, 15(3):178–196.
- Lehtinen L, Ketola K, Makela R, Mpindi JP, Viitala M, Kallioniemi O, Iljin K. 2013. High-throughput RNAi screening for novel modulators of vimentin expression identifies MTHFD2 as a regulator of breast cancer cell migration and invasion. Oncotarget, 4(1):48-63.
- Li L, Li W. 2015. Epithelial-mesenchymal transition in human cancer: comprehensive reprogramming of metabolism, epigenetics, and differentiation. Pharmacology & Therapeutics, 150:33-46.
- Liem KF Jr, Jessell TM, Briscoe J. 2000. Regulation of the neural patterning activity of sonic hedgehog by secreted BMP inhibitors expressed by notochord and somites. Development, 127(22): 4855-4866.
- Lv JF, Shim JS. 2015. Existing drugs and their application in drug discovery targeting cancer stem cells. Archives of Pharmacal Research, 38(9):1617–1626.
- Mani SA, Yang J, Brooks M, Schwaninger G, Zhou A, Miura N,Kutok JL, Hartwell K, Richardson AL, Weinberg RA. 2007.Mesenchyme Forkhead 1(FOXC2) plays a key role in

- metastasis and is associated with aggressive basal-like breast cancers. Proceedings of the National Academy of Sciences of the United States of America, 104(24):10069–10074.
- Martin FT, Dwyer RM, Kelly J, Khan S, Murphy JM, Curran C, Miller N, Hennessy E, Dockery P, Barry FP, O'Brien T, Kerin MJ. 2010. Potential role of mesenchymal stem cells (MSCs) in the breast tumour microenvironment: stimulation of epithelial to mesenchymal transition (EMT). Breast Cancer Research and Treatment, 124(2):317-326.
- Medici D, Hay ED, Olsen BR. 2008. Snail and Slug promote epithelial-mesenchymal transition through the beta-catenin-T-cell factor-4-dependent expression of transforming growth factor-beta3. Molecular Biology of the Cell, 19(11):4875–4887.
- Meidhof S, Brabletz S, Lehmann W, Preca BT, Mock K, Ruh M, Schüler J, Berthold M, Weber A, Burk U, Lübbert M, Puhr M, Culig Z, Wellner U, Keck T, Bronsert P, Küsters S, Hopt UT, Stemmler MP, Brabletz T. 2015. ZEB1-associated drug resistance in cancer cells is reversed by the class I HDAC inhibitor mocetinostat. EMBO Molecular Medicine, 7(6): 831–847.
- Namba T, Kodama R, Moritomo S, Hoshino T, Mizushima T. 2015. Zidovudine, an anti-viral drug, resensitizes gemcitabine-resistant pancreatic cancer cells to gemcitabine by inhibition of the Akt-GSK3 beta-Snail pathway. Cell Death and Disease, 6: e1795.
- Niessen K, Fu Y, Chang L, Hoodless PA, McFadden D, Karsan A. 2008. Slug is a direct Notch target required for initiation of cardiac cushion cellularization. Journal of Cell Biology, 182(2):315–325
- Patel NA, Patel PS, Vora HH. 2015. Role of PRL-3, Snail, Cytokeratin and Vimentin expression in epithelial mesenchymal transition in breast carcinoma. Breast Disease. 35(2): 113-127.
- Phua YL, Martel N, Pennisi DJ, Little MH, Wilkinson L. 2013. Distinct sites of renal fibrosis in Crim1 mutant mice arise from multiple cellular origins. The Journal of Pathology, 229(5):685-696.
- Pickup M, Novitskiy S, Moses HL. 2013. The roles of TGFβ in the tumour microenvironment. Nature Reviews Cancer, 13(11):788-799.
- Potenta S, Zeisberg E, Kalluri R. 2008. The role of endothelial-to-mesenchymal transition in cancer progression. British Journal of Cancer, 99(9):1375-1379.
- Ren T, Zhang W, Liu X, Zhao H, Zhang J, Zhang J, Li X, Zhang Y, Bu X, Shi M, Yao L, Su J. 2014. Discoidin

- domain receptor 2 (DDR2) promotes breast cancer cell metastasis and the mechanism implicates epithelial-mesenchymal transition program under hypoxia. The Journal of Pathology, 234(4):526-537.
- Singhai R, Patil VW, Jaiswal SR, Patil SD, Tayade MB, Patil AV. 2011. E-Cadherin as a diagnostic biomarker in breast cancer. North American Journal of Medical Sciences 3(5):227-233.
- Skromne I, Stern CD. 2001. Interactions between Wnt and Vg1 signalling pathways initiate primitive streak formation in the chick embryo. Development, 128(15):2915–2927.
- Smit MA, Peeper DS. 2008. Deregulating EMT and senescence: double impact by a single twist. Cancer Cell, 14(1):5-7.
- Strutz F, Zeisberg M, Ziyadeh FN, Yang CQ, Kalluri R, Müller GA, Neilson EG. 2002. Role of basic fibroblast growth factor-2 in epithelial-mesenchymal transformation. Kidney International, 61(5):1714-1728.
- Taki M, Verschueren K, Yokoyama K, Nagayama M, Kamata N. 2006. Involvement of Ets-1 transcription factor in inducing matrix metalloproteinase-2 expression by epithelialmesenchymal transition in human squamous carcinoma cells. International Journal of Oncology, 28(2):487–496.
- Thiery JP, Acloque H, Huang RY, Nieto MA. 2009. Epithelial-mesenchymal transitions in development and disease. Cell, 139(5):871–890.
- Thiery JP, Sleeman JP. 2006. Complex networks orchestrate epithelial-mesenchymal transitions. Nature Reviews Molecular Cell Biology, 7(2):131–142.
- Thiery JP. 2002. Epithelial-mesenchymal transition in tumour progression. Nature Reviews Cancer, 2(6):442–454.
- Thomas P, Brickman JM, Popperl H, Krumlauf R, Beddington RS. 1997. Axis duplication and anterior identity in the mouse embryo. Cold Spring Harbor Symposia on Quantitative Biology, 62:115–125.
- Thompson EW, Newgreen DF, Tarin D. 2005. Carcinoma invasion and metastasis: a role for epithelial-mesenchymal transition? Cancer Research, 65(14):5991-5995
- Thompson EW, Torri J, Sabol M, Sommers CL, Byers S, Valverius EM, Martin GR, Lippman ME, Stampfer MR, Dickson RB. 1994. Oncogene-induced basement membrane invasiveness in human mammary epithelial cells. Clinical & Experimental Metastasis, 12(3):181–194
- Toden S, Okugawa Y, Jascur T, Wodarz D, Komarova NL, Buhrmann C, Shakibaei M, Boland CR, Goel A. 2015. Curcumin mediates chemosensitization to 5-fluorouracil through miRNA-induced suppression of epithelial-tomesenchymal transition in chemoresistant colorectal cancer. Carcinogenesis, 36(3):355–367.

- Tse JC, Kalluri R. 2007. Mechanisms of metastasis: epithelial-to-mesenchymal transition and contribution of tumor microenvironment. Journal of Cellular Biochemistry, 101(4):816–829.
- Tse JC, Kalluri R. 2007. Mechanisms of metastasis: epithelial-to-mesenchymal transition and contribution of tumor microenvironment. Journal of Cellular Biochemistry, 101(1):816-829.
- Vaittinen S, Lukka R, Sahlgren C, Hurme T, Rantanen J, Lendahl U, Eriksson JE, Kalimo H. 2001. The expression of intermediate filament protein nestin as related to vimentin and desmin in regenerating skeletal muscle. Journal of Neuropathology & Experimental Neurology, 60(6):588-597.
- Vazquez-Martin A, Oliveras-Ferraros C, Cufi S, Del Barco S, Martin-Castillo B, Menendez JA. 2010. Metformin regulates breast cancer stem cell ontogeny by transcriptional regulation of the epithelial-mesenchymal transition (EMT) status. Cell Cycle, 9(18), 3807–3814.
- Vicovac L, Aplin JD. 1996. Epithelial mesenchymal transition during trophoblast differentiation. Acta Anatomica (Basel), 156(3):202–216.
- Wang Y, Zhou BP. 2011. Epithelial-mesenchymal transition in breast cancer progression and metastasis. Chinese Journal of Cancer, 30(9):603-611.
- Xue C, Plieth D, Venkov C, Xu C, Neilson EG. 2003. The gatekeeper effect of epithelial mesenchymal transition regulates the frequency of breast cancer metastasis. Cancer Research, 63(12):3386–3394.
- Yu Y, Xiao CH, Tan LD, Wang QS, Li XQ, Feng YM. 2014. Cancer-associated fibroblasts induce epithelial-mesenchymal transition of breast cancer cells through paracrine TGF-β signaling. British Journal of Cancer, 110(3):724-732.
- Zeisberg EM, Tarnavski O, Zeisberg M, Dorfman AL, McMullen JR, Gustafsson E, Chandraker A, Yuan X, Pu WT, Roberts AB, Neilson EG, Sayegh MH, Izumo S, Kalluri R. 2007. Endothelial-to-mesenchymal transition contributes to cardiac fibrosis. Nature. Medicine, 13(8):952–961.
- Zeisberg M, Neilson EG. 2009. Biomarkers for epithelial-mesenchymal transitions. Journal of Clinical Investigation, 119(6):1429–1437.
- Zeisberg M, Yang C, Martino M, Duncan MB, Rieder F, Tanjore H, Kalluri R. 2007. Fibroblasts derive from hepatocytes in liver fibrosis via epithelial to mesenchymal transition. Journal of Biological

- Chemistry, 282(32):23337-23347.
- Zhao Z, Cheng X, Wang Y, Han R, Li L, Xiang T, He L, Long H, Zhu B, He Y. 2014. Metformin inhibits the IL-6-induced epithelial-mesenchymal transition and lung adenocarcinoma growth and metastasis. PLoS ONE, 9: e95884.
- Zhou Y, Liang C, Xue F, Chen W, Zhi X, Feng X, Bai X, Liang T. 2015. Salinomycin decreases doxorubicin resistance in hepatocellular carcinoma cells by inhibiting the beta-catenin/TCF complex association via FOXO3a activation. Oncotarget, 6(12):10350-10365.