

Research Article**Scientific coformer screening, preparation and evaluation of Dabigatran Etexilate Mesylate Cocystal****Shankar Pol^{1*}, Rajesh Nawale², Prashant Puranik³, Hrutuja Chalak⁴, Harsha Pol⁵**¹Department of Pharmacy, Research Scholar, YB Chavan College of Pharmacy, Dr. BAMU, Aurangabad - 431001, Maharashtra, India.²Department of Pharmacology, Faculty of Pharmacy, Government College of Pharmacy, Dr. BAMU, Aurangabad - 431005, Maharashtra, India.³Department of Pharmaceutics, Faculty of Pharmacy, University Department of Pharmaceutical Sciences, RTMNU, Nagpur-440033, Maharashtra, India.^{4,5}ZIM Laboratories Limited, Nagpur- 441501, Maharashtra, India.

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Abstract

Objective: This present study aims to screening of pharmaceutical cocystal of Dabigatran Etexilate Mesylate (DEM) and coformers using different methods. Further the preparation and evaluation of DEM-coformer cocystal and study the effect of cocrystallization on stability of DEM. **Material and Methods:** The coformers for DEM were screened using Hansen Fedor's solubility parameter. Compatibility study between DEM and coformer was performed. Coformer and cocrystallization method were selected based upon impurity level determination by RP-HPLC method during screening of coformers and cocrystallization methods. Further cocrystallization between selected coformer and DEM was confirmed using molecular docking study. The DEM-coformer cocystal prepared using neat grinding method. The cocystals produced were characterized using Differential Scanning Calorimetry (DSC), Fourier Transform Infrared (FTIR) spectroscopy and impurity level determination. **Results:** Cocystal of DEM with amino acid was successfully prepared by neat grinding method. The formation of cocystals of DEM with amino acid was evaluated by DSC, IR and impurity level determination by RP-HPLC method. **Conclusion:** The DEM-Leucine cocystal exhibit enhanced stability as compared to other cocystal. This study confirms that selection of proper coformer is very crucial step in preparation of stable, good cocystal. Based upon above study and results it revealed that cocrystallization offers a valuable way to improve the physicochemical properties of the API.

Keywords: Pharmaceutical Cocystal, Dabigatran Etexilate Mesylate, Coformer, Hansen Fedor's solubility parameter

Introduction

Pharmaceutical cocrystallization is a reliable method to modify and improve physical and technical properties of drugs such as physical and chemical stability, solubility, dissolution rate, hygroscopicity and compressibility without altering their pharmacological activity or behaviour (Almarsson and Zaworotko, 2004; Schultheiss and Newman, 2009). Pharmaceutical cocystals provide an alternative to chemical modification of the drug substance as well as established salt,

amorphous, solvate, polymorphic drug forms and inclusion complexes, all of which have limitations in their utility (Mathew, 2009; Srikanth et al., 2010). Cocystal formation depends on the functional groups between API and coformer, to allow for the occurrence of hydrogen bonds or other forms of solid interaction (Namara et al., 2006). Pharmaceutical cocystal is a solid form built using synthon-based design, where the API and cocystal former molecules (coformer) connected through strong supramolecular synthons (Desiraju, 1995). Additionally Pharmaceutical cocystals are the crystalline materials comprised of two or more compound both of which are solids at room temperature, bond together in a crystal lattice through non-covalent intermolecular interactions, often including hydrogen bonding (Mohammad et al., 2011).

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Cofomers were chosen as they have some pharmaceutical relevance, i.e. APIs or biological building blocks. The other criterion was that they needed to be computationally feasible, i.e. have limited flexibility and contain only carbon, hydrogen, oxygen and nitrogen in common functional groups. A further limitation was that the compounds could be safely handled in the available facilities. We used amino acids as cofomers to obtain cocrystals of DEM. Amino acids are natural to the body, which makes them perfect candidates for cocrystallization of DEM.

Dabigatran Etxilate Mesylate structure consists of one benzimidazole, one pyridine, three carbonyl groups, four aliphatic nitrogen atoms and three aromatic nitrogen atoms. DEM molecule has three hydrogen bond donors as well as eleven hydrogen bond acceptors due to aromatic nitrogen (N_{arom}) in imidazole ring, pyridine ring and carbonyl groups and significant conformational flexibility. Caffeine and theophylline are examples of APIs that have imidazole ring. Some co-crystals between caffeine and some carboxylic acids, namely carboxylic acids (Bucar et al., 2009), oxalic acid (Trask et al., 2005) and glutaric acid (Abourahma et al., 2012) have been studied. As well as caffeine, theophylline also formed co-crystal with oxalic acid (Zhang and Rasmuson, 2012), benzoic acid (Heiden et al., 2012) and salicylic acid (Namara et al., 2006). The chemical structures of DEM and Amino acids shown in figure 1 and 2.

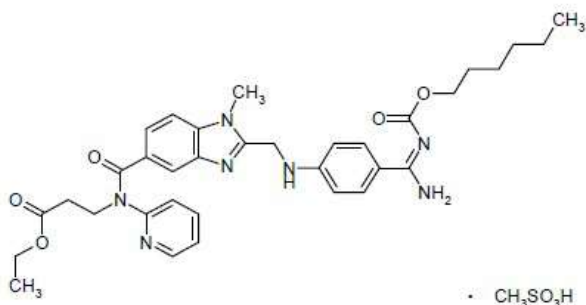


Figure 1. Dabigatran Etxilate Mesylate

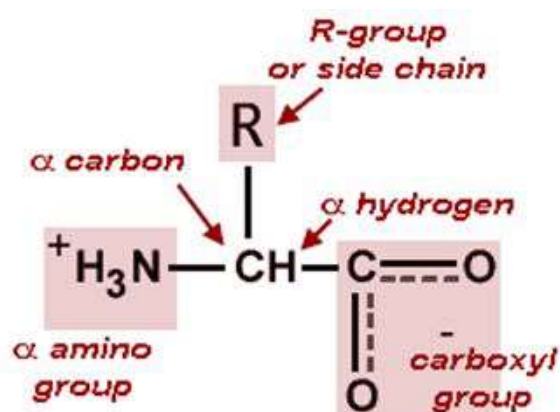


Figure 2. Amino Acid

Dabigatran Etxilate Mesylate is a BCS Class II (low solubility / high permeability) drug substance. The absolute bioavailability of Dabigatran following oral administration of Dabigatran Etxilate Mesylate is approximately 3 to 7%. Dabigatran base molecule has sub-therapeutic bioavailability and hence it is converted into prodrug i.e. Dabigatran Etxilate which leads improves bioavailability of Dabigatran, additionally further to improvement in solubility and other properties, salt form of Dabigatran Etxilate is prepared i.e. Dabigatran Etxilate Mesylate. Dabigatran Etxilate Mesylate is absorbed as the Dabigatran Etxilate ester. The ester is then hydrolyzed, forming Dabigatran, the active moiety. Dabigatran Etxilate Mesylate and its acyl glucuronides are competitive, direct thrombin inhibitors. Because thrombin (serine protease) enables the conversion of fibrinogen into fibrin during the coagulation cascade, its inhibition prevents the development of a thrombus. Both free and clot-bound thrombin and thrombin-induced platelet aggregation are inhibited by the active moieties. Dabigatran Etxilate Mesylate is long waited oral anti-coagulants after warfarin. Dabigatran Etxilate Mesylate is currently marketed as PRADAXA as oral immediate release capsule. Dabigatran Etxilate Mesylate is highly moisture sensitive, less stable and less soluble.

The aim of the study was to screening, prepare and evaluation of cocrystals of Dabigatran Etxilate Mesylate with different cofomers. For the said purpose, first Hansen Solubility Parameters (HSPs) approach was used by application of Fedor's group substitution method to investigate the miscibility of a drug and cofomer. To predict the miscibility of a drug with excipients in different formulation like solid dispersions, HSPs have been widely used (Greenhalgh et al., 1999). HSPs could predict the compatibility of pharmaceutical materials and for pre-formulation and formulation development. The HSPs of the cofomers and Dabigatran were calculated using Fedor's group substitution method. Based on the HSPs calculations, cofomers were selected for cocrystallization with Dabigatran Etxilate Mesylate (Mohammad et al., 2011). The absolute bioavailability of dabigatran following oral administration of Dabigatran Etxilate Mesylate is approximately 3 to 7%. After oral administration of radiolabeled Dabigatran, 7% of radioactivity is recovered in urine and 86% in feces (Boehringer Ingelheim, 2010). Failure to respond to Dabigatran therapy may arise due to poor aqueous solubility, pH dependent solubility, and highly sensitive to degradation in presence of moisture, acidic conditions. Previously to improve the solubility, stability, the preparation of a solid dispersion with

cyclodextrins, or/ providing acidic microenvironment to Dabigatran Etexilate by modification of formulation strategy like coating, separation Dabigatran Etexilate and Tartaric acid within same formulation was reported (Ulrich and Norbert, 2003; Boeck, 2013).

Thus, the development of the readily absorbed, stable oral anticoagulant drug is an unmet need for treatment to reduce the risk of stroke and systemic embolism in patients with non-valvular atrial fibrillation disease.

Overall study of Dabigatran Etexilate Mesylate, coformer, cocrystallization technology and most important unmet need gives idea to applicant that DEM has a higher chance to form a cocrystal with amino acids. It is very essential to prepare Dabigatran Etexilaaate Mesylate – Amino Acid (DEM-AA) cocrystal, so it can improve stability and/or solubility of DEM. The purpose of this study was to prepare and characterize DEM-AA cocrystal.

Cocrystal screening is an experimental process to determine that a particular coformer candidate is able to cocrystallise with a targeted API. In Cocrystal screening proper cofomers could be selected to do scale up experiments.

Cocrystals are miscible systems at a molecular level, hence the proposition was brought in that miscibility could be a good indication tool of cocrystal formation between two molecules in the solid state, which would help researchers to avoid going through exhausting cocrystal screening studies. Hence in the present study, the selection of cocrystal system is based on Hansen solubility parameter (HSP) calculated by Fedor's group substitution method for drug and all CCFs.

Materials and Methods

Materials

DEM is provided by ZIM laboratories Limited. All amino acid cofomers purchased from central scientific company. All required solvents and excipients provided by ZIM laboratories Limited.

Selection of cofomers

At Initial level, cofomers study based upon literature, we selected 63 cofomers for further detailed screening and evaluation based upon coformer compatibility with DEM, miscibility parameter. We selected 25 cofomers based on incompatibility of DEM with acidic nature excipients (Ulrich and Norbert, 2003). Further, selection of cofomers based upon calculation of solubility parameter.

Solubility parameter

The ideal pharmaceutical formulations have optimised solubility and stability of drug molecule. Solubility of drug is important parameter where the solubility plays a vital role in the development of pharmaceutical formulation with optimized

physical properties for readily bioavailability of drug. In general, solubility parameters are termed as cohesion energy parameters and derive from the energy needed to convert a liquid phase to a gas phase. The energy of vaporization is direct measures of the total (cohesive) energy present in the liquid's molecules together. The term cohesion energy parameter is more appropriately used when referred to surface phenomena.

$$c = \frac{\Delta H - RT}{V_m} \dots\dots\dots (1)$$

Where,

c=Cohesive energy density,

H=Heat of vaporization,

R=Gas constant,

T=Temperature,

V_m=Molar volume.

The cohesive energy density (CED) of a liquid phase is a numerical value, indicating the energy of vaporization in calories per cubic centimeter, and is a directly reflecting to degree of van der Waals forces holding the molecules of the liquid together. The correlation between vaporization and van der Waals forces helpful in transforming into a correlation between vaporization and solubility behaviour, because the same intermolecular attractive forces have to be overcome to vaporize a liquid as to dissolve it. When intermolecular attractive forces are similar, it means such material have higher solubility within each other. It is experienced in previous research and can expect that material with similar cohesive energy density (CED) would be miscible within each other (Gaikwad et al., 2017; Hansen, 2007).

HSPs calculations of Dabigatran

The concept of a solubility parameter (δ) was introduced by Hildebrand and Scott, who proposed that materials with similar δ values would be miscible (Hansen, 2007). The HSP model, which was developed later, is based on the concept of dividing the total cohesive energy into individual components (dispersion, polar and hydrogen bonding). In pharmaceutical sciences, HSPs have been used to predict the miscibility of a drug with excipients/carriers in solid dispersions, cocrystallization, complex formation, drug-excipient compatability study, preformulation study and many other applications (Hansen, 1967; Etter, 1990; Hancock et al., 1997; Greenhalgh et al., 1999; Mohammad et al., 2011).

Hildebrand et al. named the energy of vaporization per unit volume as the CED.

$$\delta = (CED)^{0.5} = (\Delta E/V)^{0.5} \dots\dots\dots (2)$$

Where,

E is the energy of vaporization

V is the molar volume

Hansen assumed that total cohesion energy is the sum of dispersion E_D , polar E_P , and hydrogen bond energy E_H .

$$E_T = E_D + E_P + E_H \dots\dots\dots (3)$$

And by dividing both sides of the equation by molar volume V, we will have the total Hansen solubility parameter or Hildebrand solubility parameter δ_T .

$$\delta_T^2 = \delta_D^2 + \delta_P^2 + \delta_H^2 \dots\dots\dots (4)$$

Where,

δ_T =Total solubility parameter

δ_D =Dispersion interactive (London) force

δ_P =Permanent dipoles in interacting molecules, called dipole-dipole interactive forces

δ_H =Hydrogen-bonding force

The common used units for δ in literatures are (J/m³), MPa, or (cal/cm³), where one (cal/cm³) is equivalent to 2.0421 MPa or (J/m³).

Solubility parameters for dry solutes may be obtained by Fedor's group substitution method. The basic steps in Fedor's method are to open the rings and treat the resultant structure as an open-chain compound. Then, the approximate substituent constants are applied. These are summed, and the solubility parameter calculated as square root of the sum of energy of mixing substituent constants divided by the sum of molar volume substituent constants (Fedors, 1974).

The Fedor's group substitution method is used for theoretical calculation which helps for the selection of cofomer which is compatible with drug. The Fedor's group substitution method reduces practical work by predicting whether the molecular complex is formed or not. The Fedor's method calculation is based on the attachment of atom or molecules from the structure. This method is used for theoretical calculation of solubility. The theoretical prediction or possibility of cocrystal formation predicted by Krevelen's $\Delta \leq 5$ MP and Greenhalgh $\Delta \leq 7$ MP respectively (Fedors, 1974; Greenhalgh et al., 1999; Savova et al., 2007). Compounds with similar δ values are likely to be miscible (Ozdemir and Guner, 2007), hence we selected closer δ values i.e. $\Delta \leq 2$ MP for present study.

Fedor's group substitution method

Fedor's proposed a method of determining solubility parameter without using the density value of the compound. This method is supposed to be better than Small's method for two reasons: The contributions of much larger number of functional groups have been evaluated, and the method requires only the knowledge of structural formula of the compound (Rathi, 2010; Gaikwad et al., 2017). The following equation is used for directly determining solubility parameter:

$$\delta_2 = \sqrt{\frac{\sum \Delta u}{\Delta V}} \dots\dots\dots (5)$$

Where, Δu and ΔV are the cohesive energy per mole and molar volume respectively.

Based on Fedor's Substitution constants,

$$\delta_2 = \sqrt{\frac{\sum \Delta u}{\Delta V}} = 12.06 \text{ H}$$

Total solubility parameter ($\Delta \delta t$) between the drug and the cofomers can be applied as a tool to predict the miscibility of two compounds.

$$\Delta \delta t = |\delta t_2 - \delta t_1| \dots\dots\dots (6)$$

Where t1 and t2 are cofomers and drug respectively and compounds with $\Delta \delta t < 7 \text{ MP}^{0.5}$ are miscible with each other (Mohammad et al., 2011). The theoretical prediction of cocrystal formation is shown in Table 2. Compounds with similar δ values are likely to be miscible (Ozdemir and Guner, 2007), hence we selected closer δ values i.e. $\Delta \leq 2$ MP for present study.

Cofomer screening for preparation of DEM Cocrystals

We did compatibility study of selected 13 cofomers after application of solubility parameter criteria based upon Fedor's group substitution method. We selected 4 cofomers based upon compatibility study results. Additionally computer docking study performed for possibility and confirmation of formation of cocrystal between DEM and four selected cofomers. Further we used selected four cofomers for cocrystallization method.

Compatibility study

Study of drug-excipient interactions is an important step in the preformulation stage of development of a dosage form. Excipients can influence the stability and bioavailability of

formulations by physical and chemical interactions with active pharmaceutical ingredient (Jackson et al., 2000; Rowe et al., 2009). These interactions may lead to change in the physicochemical properties of the drug and generation of degradation products (Chadha and Bhandari, 2014). It is therefore important to conduct drug-excipient compatibility studies to know possible interactions for the development of a stable and effective pharmaceutical formulation.

We stored DEM-coformers mixtures in two ratio (1:1 and 1:2) at 40°C / 75% RH for 1 month and evaluated RS-DEM by RP-HPLC analytical method. Chromatographic analysis was performed on Princeton SPHER-100 C18 (250 X 4.6 mm, 5 µm) HPLC column, maintained at 30°C column temperature, 6°C sample tray temperature and detection monitored at 225 nm. The mobile phase consisted of Acetonitrile: Phosphate Buffer (pH 2.5) (33:67 V/V). The flow rate was maintained at 1.0 ml/min. We selected four coformers for further preparation of cocrystal and selection of method of cocrystallization on the basis lowest impurity level after RS-DEM analysis.

Molecular Docking

Molecular docking is effective approach for computer aided structure-based drug discovery which predicts the possibility of binding and preferred orientation of one molecule to a second when bound to each other to form a new complex (Lengauer and Rarey, 1996). Information of the binding and preferred orientation is used to predict the strength of association or

binding affinity between two molecules using scoring functions which is helpful for designing experiments with correct view and approach (Gaba et al., 2010; Meng et al., 2011). We confirmed the possibility of formation of cocrystal of above selected four coformers based on solubility parameter, molecular docking study and compatibility study.

Preparation of DEM cocrystals

Synthesis of cocrystals of DEM was carried out using various methods like grinding method, solvent drop grinding method, grinding with sonication method and solvent drop grinding with sonication method. The type and amount of solvent and ratio of the API with Coformer is an important variable in cocrystallization. We selected methanol as solvent on the basis of DEM, coformer solubility and literature survey.

a) Neat grinding method

The accurately weighed quantity of DEM and coformers in 1:1, 1:2 molar ratio was grounded in mortar and pestle for 30 min, the cocrystals obtained was collected and stored in desiccator till further use.

b) Solvent-drop grinding

In the solvent-drop grinding method, DEM and coformers were weighted in 1:1, 1:2 molar ratio was grounded together with addition of 3 to 4 ml of methanol which is added in drop wise manner. The mixture was grounded in mortar and pestle for 30 min, the cocrystals obtained was collected and stored in desiccator till further use.

Table 1. Calculation of solubility parameter of DEM by Fedor's group substitution method

Drug DEM fragments	No. of fragment	Cohesive Energy (Cal/Mol)	Total Cohesive Energy (ΔEV) (Cal/Mol)	Molar Volume (Cm ³ /Mol)	Total Molar Volume (Vm) (Cm ³ /Mol)
- CH3	3	1125	3375	33.5	100.5
- CH2 -	9	1180	10620	16.1	144.9
- CO	1	4150	4150	10.8	10.8
- COO	2	4300	8600	18	36
- N=	2	2800	5600	5	10
- N <	2	1000	2000	-9	-18
- NH	3	2000	6000	4.5	13.5
Phenylene	1	7630	7630	52.4	52.4
6 membered ring	2	250	500	16	32
5 membered ring	1	250	250	18	18
(=) in Ring	7	400	2800	-2.2	-15.4
=CH -	7	1030	7210	13.5	94.5
= C<(Inside Ring)	5	1030	5150	-5.5	-27.5
= C<(Other than Ring)	1	1030	1030	-5.5	-5.5
Total	46		64915		446.2
δ2(Cal/cm3)^{0.5}		(64915/446.2)^{0.5}		12.06	

c) Grinding with sonocrystallization method

In the grinding with sonication method, DEM and coformer in 1:1, 1:2 molar ratio were grinded with sonication. The mixture was grounded in mortar and pestle for 30 min, the cococrystals obtained was collected and stored in desiccator till further use.

d) Solvent-drop grinding with sonocrystallization method

In the solvent-drop grinding with sonication method, DEM and coformers were weighted in 1:1, 1:2 molar ratio was grounded together with sonication, simultaneously addition of 3 to 4 ml of methanol which is added in drop wise manner. The mixture was grounded in mortar and pestle for 30 min, the cococrystals obtained was collected and stored in desiccator till further use.

Evaluation of Cococrystal formation

The prepared cococrystal in present study was primarily confirmed by comparing DSC results, FTIR results of cococrystals with DEM (pure drug) and respective coformers. Further cococrystals evaluated for one month stability (RS-DEM) study by RP-HPLC analytical method.

Infrared spectroscopy

IR spectra of samples were recorded on FTIR IRAffinity-1S

(Shimadzu). The spectra were collected over the range of 4000-700 cm^{-1} for each sample.

Differential Scanning Calorimetry

The samples were analysed by Differential Scanning Calorimeter (Shimadzu DSC-60) over the range of 50-300 $^{\circ}\text{C}$ at the rate of 20 $^{\circ}\text{C}$ per minute.

RS-DEM Stability Evaluation of Cococrystals

RS-DEM stability of cococrystals evaluated for 1 month against controlled sample of DEM by using RP-HPLC method. We used same chromatographic analysis which is used for above compatibility study.

Results and Discussion**Miscibility prediction of coformers with DEM using Fedor's group substitution method**

The selection of appropriate cococrystal formers for pharmaceutical cococrystallization of DEM was accomplished via the miscibility of a drug and CCFs (as calculated by Fedor's group substitution method). The representative example of solubility parameter calculation

Table 2. Theoretical prediction of miscibility by Fedor's method

Compound	δ value	Difference $\delta_1 - \delta_2$	$\Delta\delta$	Possibility of cococrystal formation [$\Delta\delta \leq 2\text{MP}$]
DEM (Drug)	12.06	12.06	-	-
Alanine	11.99	12.06 - 11.99	0.07	Highly Miscible, Selected
Arginine	13.66	12.06 - 13.66	1.6	Highly Miscible, Selected
Asparagine	14.21	12.06 - 14.21	2.15	Not Highly Miscible, Rejected
Aspartic acid	14.11	12.06 - 14.11	2.05	Not Highly Miscible, Rejected
Cysteine	12.87	12.06 - 12.87	0.81	Highly Miscible, Selected
Glutamine	13.73	12.06 - 13.73	1.67	Highly Miscible, Selected
Glutamic acid	13.43	12.06 - 13.43	1.37	Highly Miscible, Selected
Glycine	12.99	12.06 - 12.99	0.93	Highly Miscible, Selected
Histidine	14.96	12.06 - 14.96	2.9	Not Highly Miscible, Rejected
Isoleucine	10.67	12.06 - 10.67	1.39	Highly Miscible, Selected
Leucine	10.67	12.06 - 10.67	1.39	Highly Miscible, Selected
Lysine	11.78	12.06 - 11.78	0.28	Highly Miscible, Selected
Methionine	11.79	12.06 - 11.79	0.27	Highly Miscible, Selected
Phenylalanine	11.97	12.06 - 11.97	0.09	Highly Miscible, Selected
Proline	11.71	12.06 - 11.71	0.35	Highly Miscible, Selected
Serine	16.03	12.06 - 16.03	3.97	Not Highly Miscible, Rejected
Threonine	14.78	12.06 - 14.78	2.72	Not Highly Miscible, Rejected
Tryptophan	17.96	12.06 - 17.96	5.9	Not Highly Miscible, Rejected
Tyrosine	14.51	12.06 - 14.51	2.45	Not Highly Miscible, Rejected
Valine	10.94	12.06 - 10.94	1.12	Highly Miscible, Selected
Urea	14.36	12.06 - 14.36	2.30	Not Highly Miscible, Rejected
Glucosamine	22.51	12.06 - 22.51	10.45	Not Highly Miscible, Rejected
Xylitol	22.59	12.06 - 22.59	10.53	Not Highly Miscible, Rejected
Meglumine	20.77	12.06 - 20.77	8.71	Not Highly Miscible, Rejected
Trimethamine	18.99	12.06 - 18.99	6.93	Not Highly Miscible, Rejected

Table 3. Compatibility Study Details

Compatibility Study Details	Study Code	Ratio of DEM:Coformer	Initial Impurity	15 Days Impurity	1 Month Impurity
Drug (DEM)	D	-	0.57	1.40	1.52
DEM & Cysteine	DC1	1:1	0.66	1.25	1.52
DEM & Cysteine	DC2	1:2	0.67	1.23	1.49
DEM & Glycine	DG1	1:1	0.68	1.34	1.41
DEM & Glycine	DG2	1:2	0.71	1.21	1.42
DEM & Alanine	DAL1	1:1	0.54	0.69	0.94
DEM & Alanine	DAL2	1:2	0.50	0.78	1.08
DEM & Arginine	DAR1	1:1	0.49	0.80	2.55
DEM & Arginine	DAR2	1:2	0.58	1.21	3.19
DEM & Glutamine	DGA1	1:1	0.43	0.67	0.97
DEM & Glutamine	DGA2	1:2	0.50	0.59	1.18
DEM & Glutamic Acid	DGAA1	1:1	0.50	0.98	1.09
DEM & Glutamic Acid	DGAA2	1:2	0.53	1.15	1.24
DEM & Isoleucine	DI1	1:1	0.51	0.85	1.18
DEM & Isoleucine	DI2	1:2	0.58	0.85	1.21
DEM & Leucine	DLU1	1:1	0.53	0.79	1.18
DEM & Leucine	DLU2	1:2	0.52	0.85	1.15
DEM & Lysine	DLY1	1:1	0.68	0.69	1.07
DEM & Lysine	DLY2	1:2	0.52	0.75	1.10
DEM & Methionine	DM1	1:1	0.52	0.90	1.24
DEM & Methionine	DM2	1:2	0.59	0.86	1.18
DEM & Phenylalanine	DPA1	1:1	0.57	1.03	1.28
DEM & Phenylalanine	DPA2	1:2	0.56	0.93	1.19
DEM & Proline	DPR1	1:1	0.58	1.11	2.05
DEM & Proline	DPR2	1:2	0.59	1.63	3.21
DEM & Valine	DV1	1:1	0.62	1.08	1.28
DEM & Valine	DV2	1:2	0.63	0.93	1.25

Table 4. Virtual screening of co-formers with using molecular docking

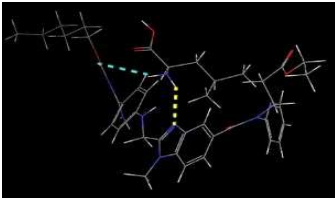
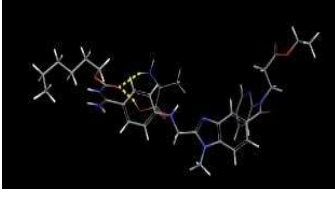
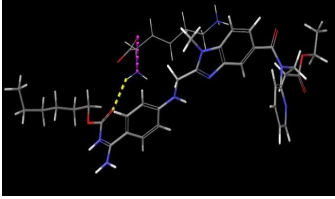
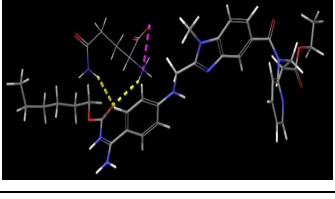
Coformer	2D structure	Interaction	Glide Score
Leucine		Yes	-1.237
Alanine		Yes	-4.14
Lysine		Yes	-1.45
Glutamine		Yes	-2.075

Table5. DSC Study Details of Cocrystal, DEM and Coformer

Cocrystal	Code	Melting Point (°C)
DEM	DEM - Drug	178.60
DEM + Alanine	DAL1 - G	174.84
DEM + Alanine	DAL1 - GS	174.61
DEM + Alanine	DAL1 - GM	172.20
DEM + Alanine	DAL1 - GSM	173.71
DEM + Glutamine	DGA1 - G	196.93
DEM + Glutamine	DGA1 - GS	196.82
DEM + Glutamine	DGA1 - GM	197.37
DEM + Glutamine	DGA1 - GSM	198.95
DEM + Leucine	DLU1 - G	174.15
DEM + Leucine	DLU1 - GS	174.37
DEM + Leucine	DLU1 - GM	172.59
DEM + Leucine	DLU1 - GSM	172.82
DEM + Lysine	DLY1 - G	173.18
DEM + Lysine	DLY1 - GS	173.78
DEM + Lysine	DLY1 - GM	174.22
DEM + Lysine	DLY1 - GSM	172.65

by Fedor's group substitution method for DEM is represented in table 1 and theoretical prediction of miscibility of DEM with coformers represented in table 2.

We selected 13 coformers based upon above coformer miscibility with DEM i.e. [$\Delta\delta \leq 2MP$] for further study.

DEM and Coformers compatibility study

All selected 13 coformers and DEM compatibility study performed in two stoichiometric ratios (1:1 and 1:2) and evaluated for 1 month by RS-DEM stability study against controlled sample of DEM by using RP-HPLC method. We found that only four coformers are more compatible with DEM. The details of 13 coformers and DEM compatibility study represented in table 3.

Molecular Docking

The molecule of DEM structure consists of two aromatic rings (imidazole and pyridine), three carbonyl groups, four aliphatic nitrogen atoms and three aromatic nitrogen atoms. DEM molecule has three hydrogen bond donors as well as eleven hydrogen bond acceptors due to aromatic nitrogen (N_{arom}) in imidazole ring, pyridine ring and carbonyl groups and significant conformational flexibility; hence it is possible to form co-crystals with certain co-formers. Co-formers chosen in this work were Leucine, glutamine, Alanine and lysine. The result of virtual screening of co-formers using molecular docking is showed in table 4.

Characterization and cocrystals detection by DSC, FTIR

The predicted cocrystals were verified experimentally by using DSC for miscibility and cocrystal detection. The signs of melting point depression were the signs of miscibility and cocrystal formation as the new phase with a new melting

Table6. RS stability study Details

Compatibility Study Details	Study Code	Initial Impurity	15 Days Impurity	1 Month Impurity
DEM	DEM	0.60	1.04	1.58
DEM Leucine Grinding	DLU G	0.75	1.19	1.61
DEM Leucine Grinding + Sonication	DLU GS	0.73	1.44	1.63
DEM Leucine Grinding + Methanol	DLU GM	0.75	1.79	1.83
DEM Leucine Grinding + Sonication + Methanol	DLU GSM	0.78	1.69	1.90
DEM Alanine Grinding	DAL G	1.66	1.74	2.07
DEM Alanine Grinding + Sonication	DAL GS	0.99	1.64	2.05
DEM Alanine Grinding + Methanol	DAL GM	0.83	1.93	2.37
DEM Alanine Grinding + Sonication + Methanol	DAL GSM	0.82	1.83	2.21
DEM Lysine Grinding	DLY G	0.90	2.38	3.86
DEM Lysine Grinding + Sonication	DLY GS	0.80	2.07	3.44
DEM Lysine Grinding + Methanol	DLY GM	0.77	1.91	2.74
DEM Lysine Grinding + Sonication + Methanol	DLY GSM	0.85	2.12	3.05
DEM Glutamine Grinding	DGA G	0.78	1.52	2.00
DEM Glutamine Grinding + Sonication	DGA GS	0.83	1.82	2.19
DEM Glutamine Grinding + Methanol	DGA GM	0.73	1.91	2.52
DEM Glutamine Grinding + Sonication + Methanol	DGA GSM	0.73	2.12	2.79

point was observed. A single endothermic sharp peak was observed for each cocrystal confirming about the new crystalline phase. Representative DSC spectra of DEM, Leucine and DEM-Leucine cocrystals with characteristic DSC endotherms at about 178.60° C, 281.61° C and 174° C respectively as shown in Figure 3, 4 and 5. However melting endotherm can be higher or lower than endotherm of API. The melting endotherms obtained from

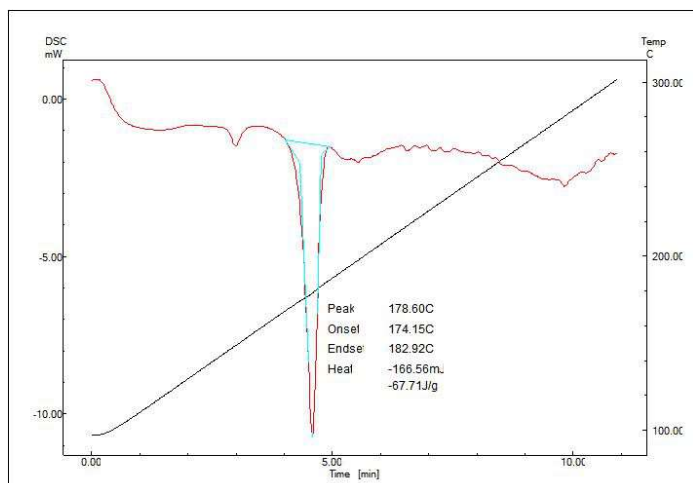


Figure3. DSC of DEM

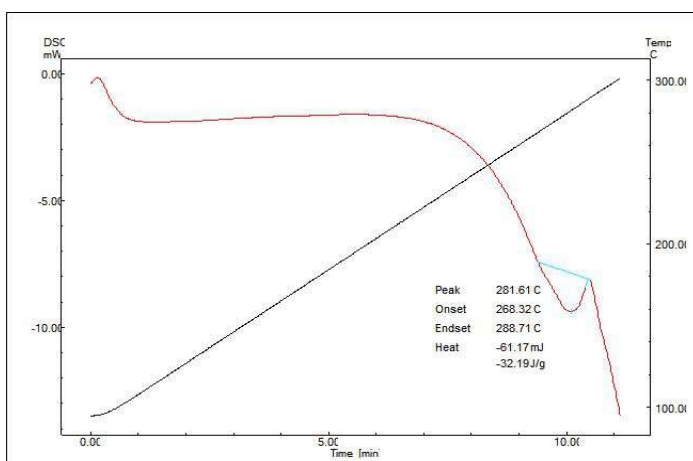


Figure4. DSC of Coformer Leucine

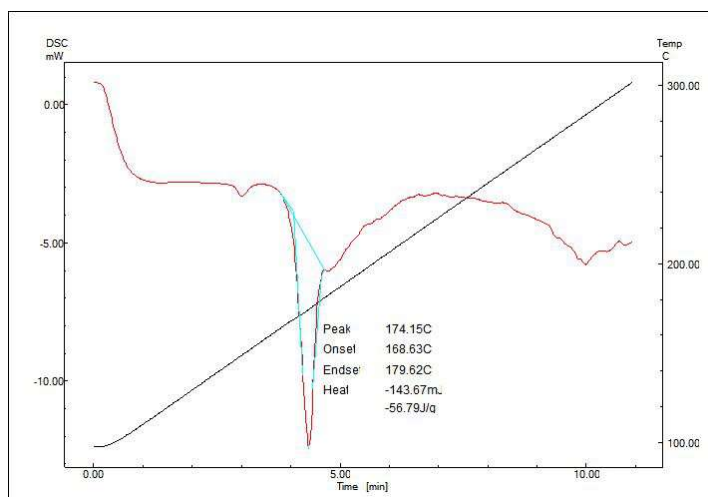


Figure5. DSC of Cocrystal DLU1 – G

DSC of all the investigated cocrystals are summarized in table 5.

Hydrogen bonding in cocrystals by FTIR spectroscopy is detected by decrease in intensity of O-H peak and appearance of low frequency broad O-H band. As seen from DEM-Leucine cocrystal, characteristic peaks at 2949, 2910, 2848, 2351 ± 5 cm^{-1} indicative of hydrogen bond formation with retention of parent drug peak. The representative FTIR spectra of DEM, Leucine and DEM-Leucine cocrystal are as shown in figure 6, 7 and 8.

After successful detection of cocrystal by DCS and FTIR, detected cocrystals were subjected to one month RS stability study against controlled sample of DEM by using RP-HPLC method and the details are represented in table 6.

DEM-Leucine cocrystal shows lowest impurity profile amongst four cocrystals of DEM & Leucine, alanine, glutamine and lysine. The summary of Cocrystal prediction and Correlation of HSP with experimental findings is represented in table 7.

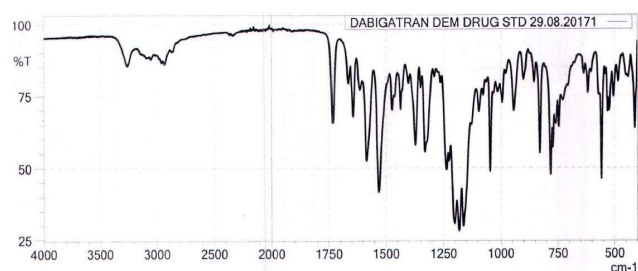


Figure6. FTIR spectra of DEM

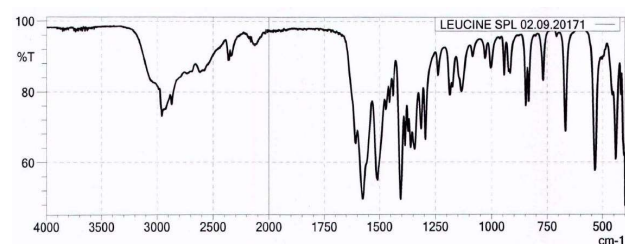


Figure 7. FTIR spectra of Leucine

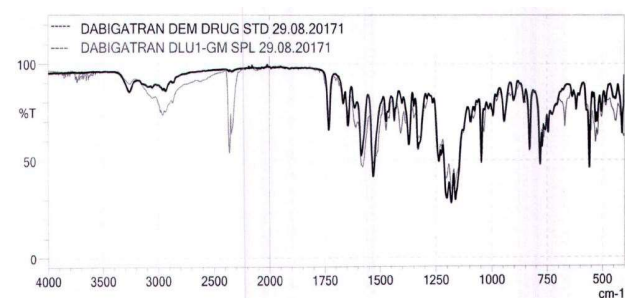


Figure8. FTIR spectra of DEM-Leucine cocrystal

Table 7. Summary of Cocrystal prediction and Correlation of HSP with experimental findings

Formulations	miscibility by HSP	DSC Melting Endotherm (°C)	IR	Stability study	Prediction and success
DEM	12.06	178.60	--	--	--
Alanine	11.99	174.84	Yes	Failed	Yes
Leucine	10.67	174.15	Yes	Passed	Yes
Lysine	11.78	173.18	Yes	Failed	Yes
Glutamine	13.73	196.93	Yes	Failed	Yes

Conclusion

The present study was aimed to investigate the use of Hansen solubility parameter in prediction of cocrystal formation between DEM and CCFs, and evaluate the stability of prepared cocrystal. The investigated approach was effective in predicting miscibility of the drug and cofomers, and 1 month stability study gives idea about formation of cocrystal and stability of prepared cocrystals. Stability of DEM cocrystals were evaluated by using RP-HPLC method.

We found that leucine –cocrystal are more stable amongst other cofomers cocrystal with DEM. Leucine is non-polar, highly hydrophobic, neutral cofomer. The hydrophobicity indices of leucine is 97% is highest amongst four amino acids. DEM is more stable with neutral cofomers as compared to acidic and basic cofomers.

The detected stable cocrystals can be further transformed to develop bioequivalent dosage as like parent form or even improved one. Future prospects of work reveal detailed evaluation of detected cocrystals for formulation, development and equivalency with marketed formulation.

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Conflicts of Interests

All authors have none to declare.

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